

INTRODUCTION

Elephants and baleen whales are massive creatures that respond to exceptionally low frequency signals. We have many examples of LF and even seismic elephant and whale vocalizations, but little direct knowledge of their hearing abilities. Playbacks and behavioral observations suggest proboscidean and mysticete hearing abilities are similarly specialized for low or even infrasonic signals (Langbauer, et al 1991; Payne, et al 1986; Wartzok and Ketten 1999).

This raises several interesting questions: First, their vocalizations are emitted and perceived in two media, air and water, which have radically different physical acoustic properties: 4.5-fold differences in sound speed; three magnitudes greater acoustic impedance:

Intensity = Sound Energy perpendicular to the direction of sound propagation

$$I = pv = p^2 / cr$$

$$I_{\text{air}} = p^2 / (340 \text{ m/sec})(0.0013 \text{ g/cc})$$

$$I_{\text{water}} = p^2 / (1530 \text{ m/sec})(1.03 \text{ g/cc})$$

$$\text{for } I_{\text{water}} = I_{\text{air}}$$

$$p_{\text{water}} = 59.7 \cdot p_{\text{air}}$$

Therefore, for elephants and whales to have the same percept, whale ears must accommodate 60-fold acoustic pressures. Further, a common concept is that the upper limit of hearing is inversely correlated with body mass, implying there should be little mid to high frequency hearing in any mysticete and little overlap in the hearing of these two taxa.

HYPOTHESIS

Whales and elephants will have common auditory structural patterns related to LF hearing but morphometric differences related to hearing in air vs water.

APPROACH

Functional analyses based on CT and histologic measurements of inner ears of Tethytheria.

MATERIALS & METHODS

PROBOSCIDAEE (7)

Loxodonta africana (African) 3 CT / 2 histology

Elephas maximus (Asian) 4 CT / 2 histology

MYSTICETI (23)

Megaptera novaeanglia (Humpback) 6 CT / 1 histology

Balaenoptera musculus (Blue) 2 CT / 1 histology

Eschrichtius robustus (Grey) 2 CT / 1 histology

Eubalaena glacialis (Right) 5 CT / 2 histology

Balaenoptera acutorostrata (Minke) 6 CT / 1 histology

Balaenoptera physalus (Finback) 2 CT / 0 histology

IMAGING

WHOI CT FACILITY <http://csi.whoi.edu>

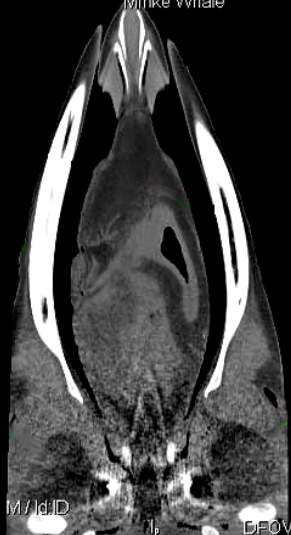
Specimens scanned on Siemens Volume Zoom as whole animals, heads, and intact temporal bones

- spiral UHR submillimeter imaging
- 0.5 mm/sec acquisitions and collimation
- UHR 90 (bone) to 40 (soft tissue) kernels
- 0.1 mm slice images
- 100 μ isotropic voxels

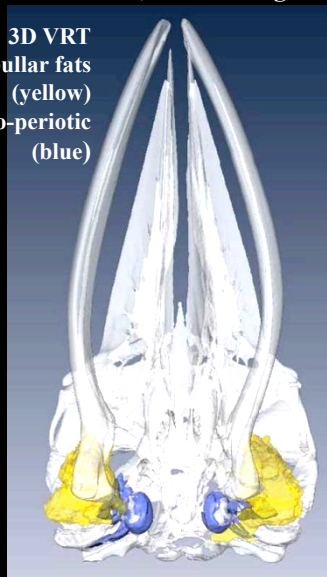
2D to 3D CT IMAGING, RECONSTRUCTION, AND MEASUREMENT (Siemens SSD -VRT, Amira)

Representative CT Scan Series for 56 cm diameter, 120 cm length intact minke whale head

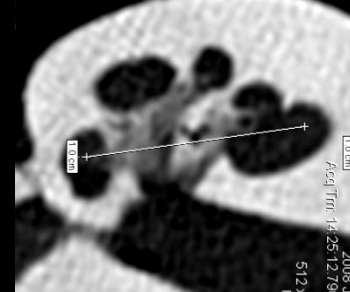
2D coronal CT Minke Whale



3D VRT peribullar fats (yellow) tympano-periotic (blue)



0.1 mm 2D modiolar section of cochlea

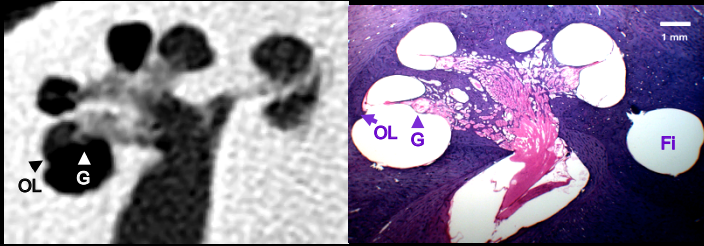


3D orthogonal view of inner ear canals



CT vs HISTOLOGY

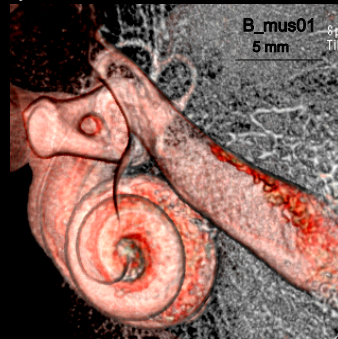
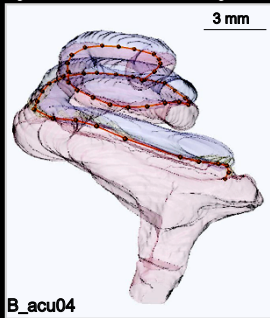
100 micron paramodiolar CT sections, 20-30 micron celloidin sections, hematoxylin-eosin stain



100 μ paramodiolar CT image (left) vs 25 μ celloidin H&E stained section (right) from the same minke whale ear. Cochlear canal dimensions are obtained from both. Histology is used for all basilar membrane measurements. Some micro-features; e.g., ganglion cell regions (G), and outer osseous lamina (OL) are visible and comparable in both the UHRCT and histology. A fiducial (Fi) marker necessary for accurate alignment of celloidin sections that is not required for 3D CT is also shown.

Cochlear Radii, Lengths, Basilar Membrane Morphometry, and Frequency Calculations (methods Ketten et al, 1998)

Multi-tissue segmentations from CT are compared with reconstructions from histology to determine shrinkage or distortion and verify calculations. The circles and subtended line represent basilar membrane positions.



Orthogonal projections are used for radii and length



Chadwick et al 2005

$$s = \int_{\theta_1}^{\theta_2} \sqrt{r^2 + \left(\frac{dr}{d\theta}\right)^2} d\theta$$

and

$$z = \sqrt{s^2 + h^2}$$

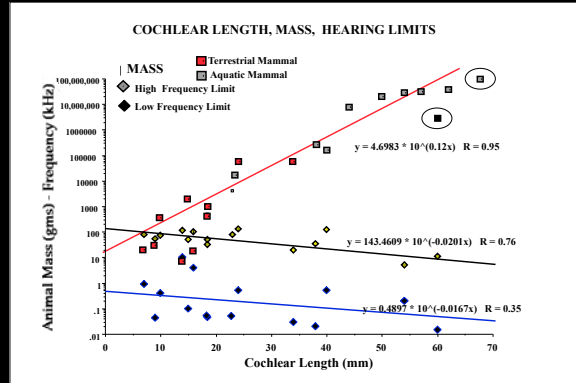
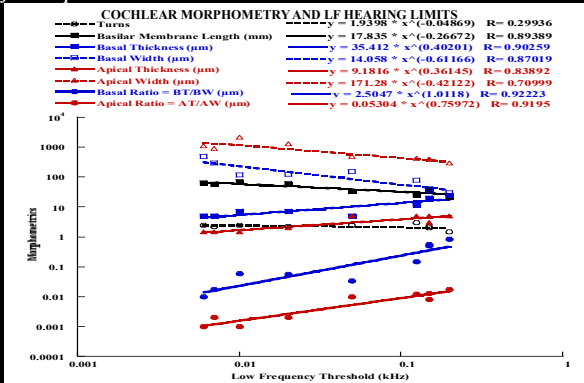
for $r = e^{a\theta}$
 $s = (r/a) \sqrt{1+a^2}$

for $r = a\theta$
 $s = (a/2) [\theta \sqrt{\theta^2+1} + \text{Ln}(\theta + \sqrt{\theta^2+1})]$

RESULTS

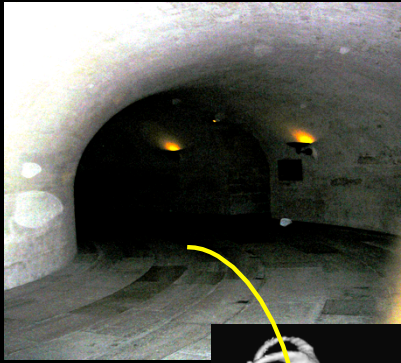
MAMMALIAN COCHLEAR MORPHOMETRY (Firbas 1972; West 1985; Echteler et al. 1994; Ketten 2000)

Species	Common Name	Hearing range (kHz)	Turns	Membrane Length (mm)	Basal Thickness (μm)	Basal Width (μm)	Apical Thickness (μm)	Apical Width (μm)
<i>Phocoena phocoena</i>	Harbor porpoise	0.3 - 180	1.5	22.5	25	30	5	290
<i>Lagenorhynchus obliquidens</i>	White-sided dolphin	0.2 - 140	2.0	33.8	20	40	3	400
<i>Tursiops truncatus</i>	Bottlenosed dolphin	0.2 - 160	2.25	38.9	20	35	5	380
<i>Balaenoptera musculus</i>	Blue whale	0.01 - 18	2.5	72.3	7	120	<2	2200
<i>Megaptera novaeangliae</i>	Humpback whale	0.02 - 30	2.25	61.65	7	125	2	1,300
<i>Loxodonta africana</i>	African elephant	0.006 - 8	2.5	65.1	5	500	2	>1100
<i>Elephas maximus</i>	Asian elephant	0.007 - 9	2.25	57.4	-	>300	-	>900
<i>Felis domesticus</i>	Cat	0.125 - 60	3.0	25.8	12	80	5	420
<i>Homo sapiens</i>	Human	0.05 - 16	2.5	33.5	5	150	5	504
<i>Spalax ehrenbergi</i>	Mole rat	0.1 - 10	3.5	13.7	9	120	18	200
<i>Pteronotus parnellii</i>	Mustached bat	16 - 110	2.75	14.3	22	50	2	110
<i>Rhinolophus ferrumequinum</i>	Horseshoe bat	7 - 90	3.25	16.1	35	80	2	150



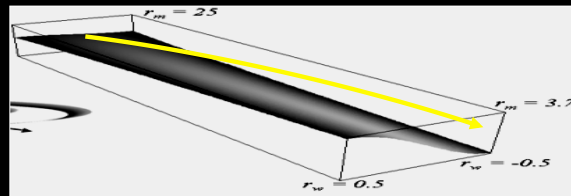
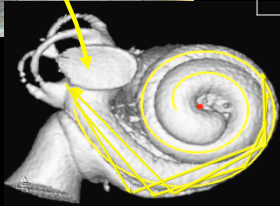
Both whales and elephants have generalist ear formats. Cochlear length is correlated with mass but is not significantly correlated with high or low frequency hearing limits. The most significant correlations for LF and HF limits of hearing are the thickness to width ratios of the basilar membrane.

Functional Effect: “Whispering Gallery” Propagation (Manoussaki et al, 2008)



Radii ratios and low-frequency threshold.							
Species	blue whale	right whale	humpback whale	bottlenose dolphin		harbor porpoise	
R_{max}/R_{min}	10.7	9.1	8.1	4.4		3.5	
LF Hz	12	15	18	150		180	
Species	african elephant	asian elephant	cow	guinea pig	man	rat	mouse
R_{max}/R_{min}	9	8.7	7.5	7.4	7	4.3	4
LF Hz	6	7	20	40	50	400	800

Broader basal curvatures and greater radii provide better propagation of LF energy to the cochlear spiral apex

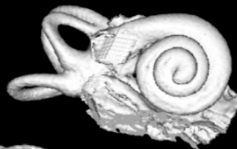


Implications: Parallel Cross-Media Strategies for LF Hearing

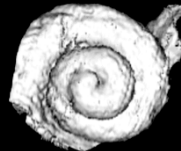
Megaptera novaeangliae



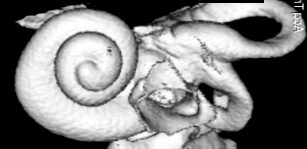
Elephas maximus



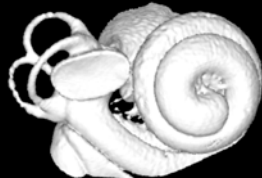
Gomphothere



Loxodonta africana



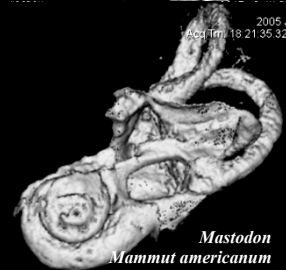
Eubalaena glacialis



Balaenoptera musculus



Mammoth primigenius



Mastodon americanum

CONCLUSIONS

Mysticetes have larger membrane gradients and are likely to have substantially greater hearing ranges than elephants (~10 vs 6-8 octaves). However, cochlear coiling and apical cochlear anatomies are similar, suggesting common mechanical adaptations for LF hearing.

LF sensitivity in both groups is strongly correlated with similar, exceptionally small apical basilar membrane ratios, which, in turn, reflect substantially lower apical stiffness as confirmed in recent point stiffness measures of some of the ears shown here (Miller et al, 2005). The ratio of minimum and maximum cochlear radii are also strongly correlated with LF thresholds and suggest there is a biologic equivalent of the classic “Whispering Gallery” effect as described by Manoussaki et al (2007).

The radii ratios are smaller in some species within each group consistent with a broadening of the basalmost regions, as exemplified by the Right Whale vs the more tightly coiled Humpback Whale shown above. Fossil proboscids show a similar division into greater radius of curvature (African elephant -Mastodon) vs lesser curvature (Asian elephant – Mammoth-Gomphothere) cochlear groups.

These results are preliminary and are hampered by sample size and preservation artifacts. They do suggest that low frequency adaptations evolved in parallel in both the mysticete and proboscidean lines over similar time scales and despite media differences. To properly address these questions, both more fossil and rare extant material should be examined with non-destructive CT techniques.

ACKNOWLEDGEMENTS

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